

Summary

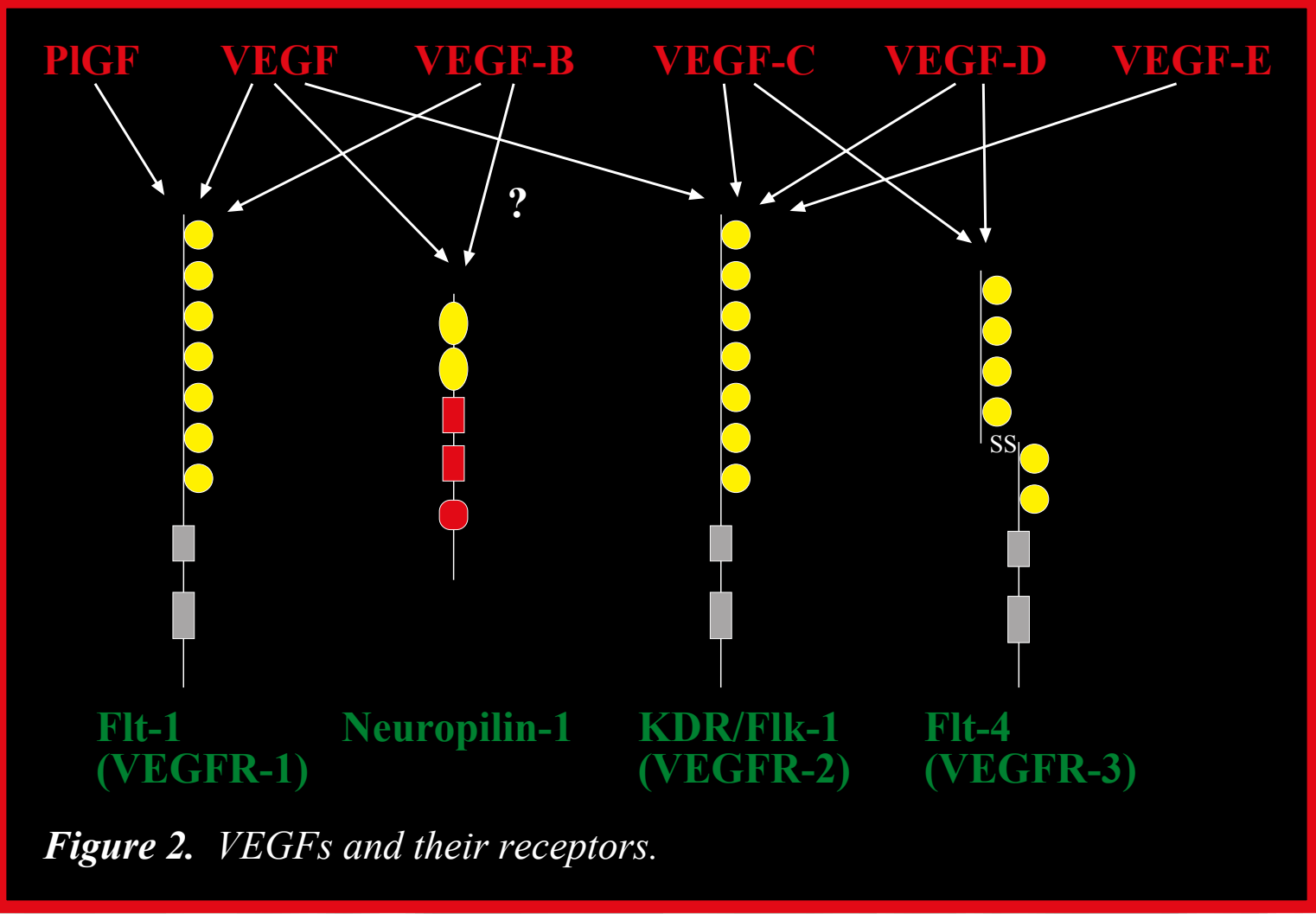
Upon reaching a size of several millimeters a developing embryo cannot meet its metabolic needs by diffusion alone. Solid tumors face the same problem, when expanding to a similar magnitude. Both embryo and tumor can only continue growing by establishing a circulatory system to supply them with oxygen and nutrients. The obvious similarity between these processes is reflected on the molecular level: Vascular endothelial growth factor (VEGF, Fig. 1) has been identified as a key regulator of blood vessel growth in both embryos and tumors. Recently we described two proteins homologous to VEGF: VEGF-C and VEGF-D. The structural similarity between VEGF-C and VEGF-D define a VEGF subfamily. Similar to Platelet-derived growth factor (PDGF) mature VEGF-C and D are generated by the cleavage of a precursor. VEGF-C plays an important role in the development of the lymphatic vascular system. The role of VEGF-D is less well defined.

Topics of our research include the further characterization of VEGF-C (biosynthesis, structure, determinants of receptor specificity) and the identification of the function of VEGF-D.

VEGFs and Their Receptors

VEGF is an important regulator of endothelial cell proliferation and migration in embryonic vasculogenesis (the in-situ differentiation of endothelial cell precursors) and angiogenesis (the growth of new blood vessels from preexisting ones).

We lately cloned two factors homologous to VEGF (JOUKOV, 1997; ACHEN, 1998), designated as VEGF-C (or *VRP*) and VEGF-D (or *FIGF*). Both VEGF-C and D are ligands for KDR (the main proliferation-inducing receptor on blood vessel endothelial cells) and Flt-4, a receptor tyrosine kinase whose expression becomes restricted to lymphatic endothelium during development (Fig. 2).



Completed Studies

**Hyperplasia of lymphatic vessels in VEGF-C transgenic mice**

VEGF-C over-expression in the basal epidermis of mice promotes the growth of lymphatic structures in the dermis, including large vessel lacunae histologically similar to lymphangioma (Figs. 3, 4, 5; JELTSCH, 1997).

**Angiogenesis and lymphangiogenesis in the chorio-allantoic membrane (CAM)**

Due to the accessibility and regularity of its vascular system the CAM has been used for many years to evaluate potential angiogenic factors (Fig. 6). Less well documented is, that the CAM is drained by a dense network of lymphatic vessels.

The effects of VEGF, PlGF, VEGF<sub>x</sub>PlGF heterodimers and VEGF-C on the differentiated avian CAM were studied (Figs. 7, 8). Only VEGF-C is chemoattractive for lymphatic endothelial cells, induces their proliferation and the development of new lymphatic sinuses (OH, 1997).

**VEGF-D is a ligand of KDR/Flk-1 and Flt-4**

VEGF-D was identified by computer-based homology search. It is most closely related to VEGF-C (Fig. 9). VEGF-D activates KDR and Flt-4. A short form of VEGF-D demonstrated receptor-binding resides in the VEGF homology domain which corresponds to the mature form of VEGF-C (Fig. 11; ACHEN, 1998).

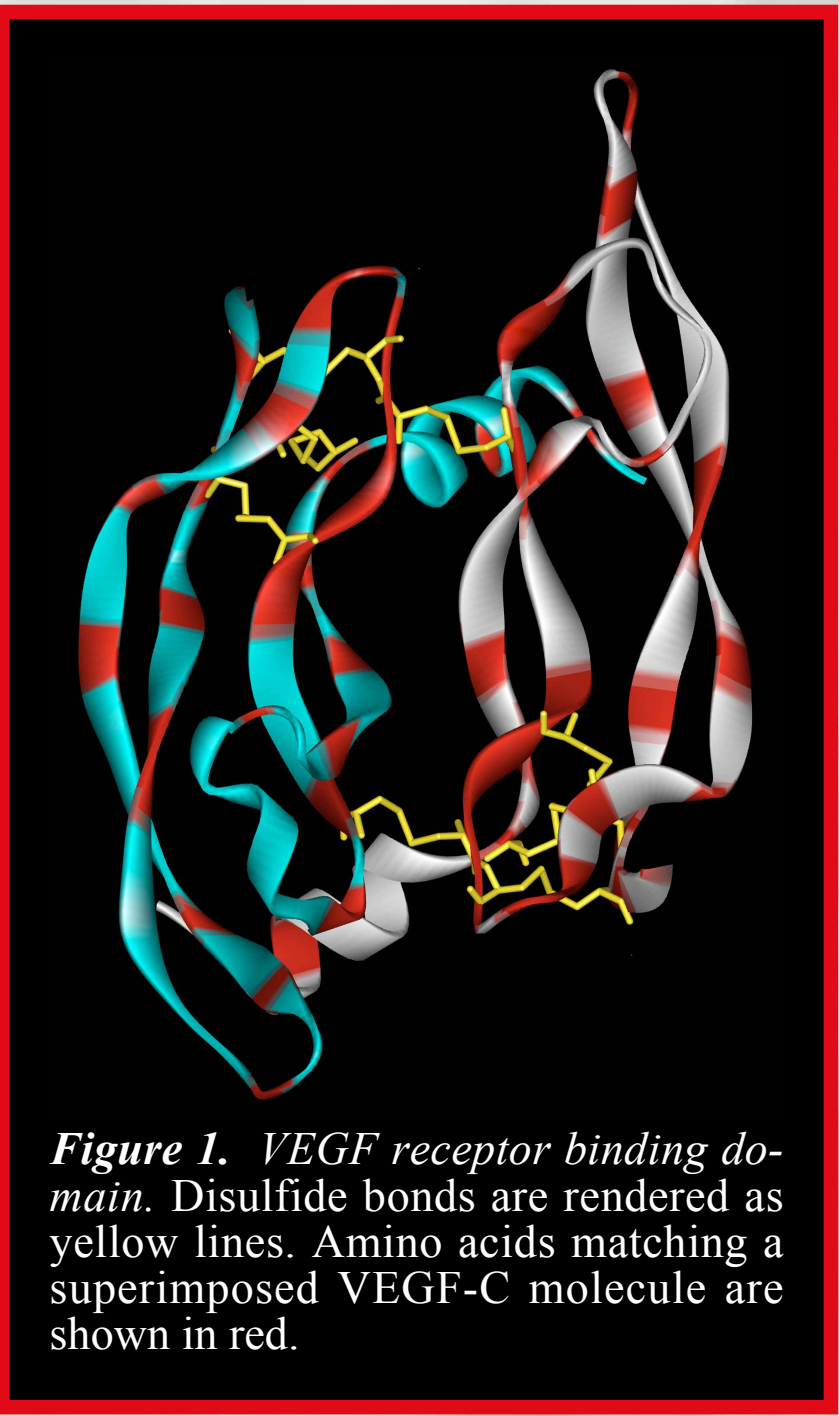
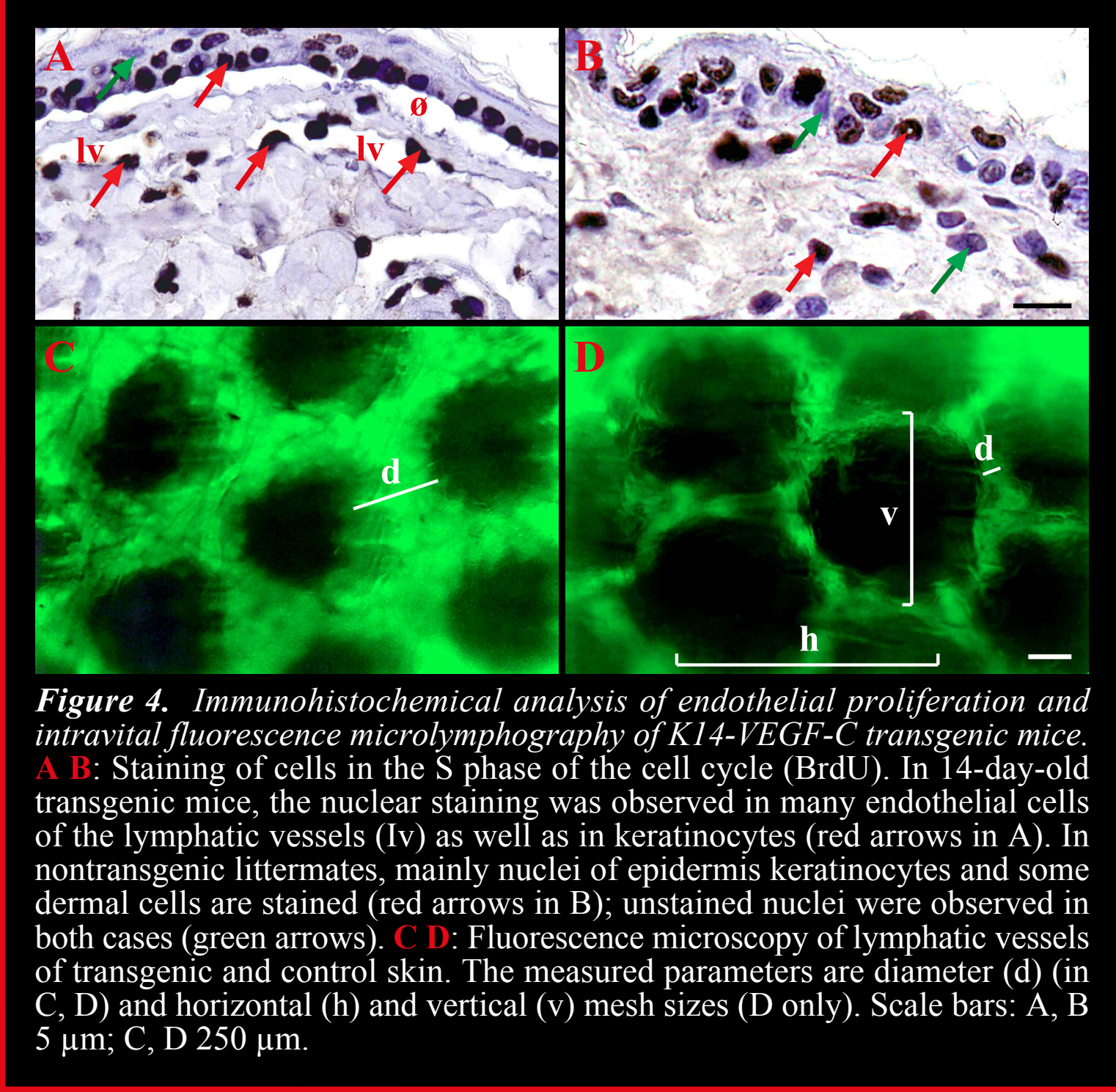


Figure 1. VEGF receptor binding domain. Disulfide bonds are rendered as yellow lines. Amino acids matching a superimposed VEGF-C molecule are shown in red.

VEGF-D Function

**Transgenic mice with tissue-specific overexpression of VEGF-D**

To identify the function of VEGF-D we are establishing animal models with tissue-specific overexpression of VEGF-D analogous to the existing VEGF-C models using the keratin 14 and the rat insulin promoter.

Recombinant Growth Factors

**Baculoviral and E.coli proteins**

Recombinant VEGF-C produced in the baculovirus system has been used in several studies (PEPPER, 1998; OH, 1997; JELTSCH, 1997; KUKK, 1996). Presently VEGF-C and VEGF-D are produced in E.coli for the production of monoclonal antibodies, in-vivo assays and crystallization purposes.

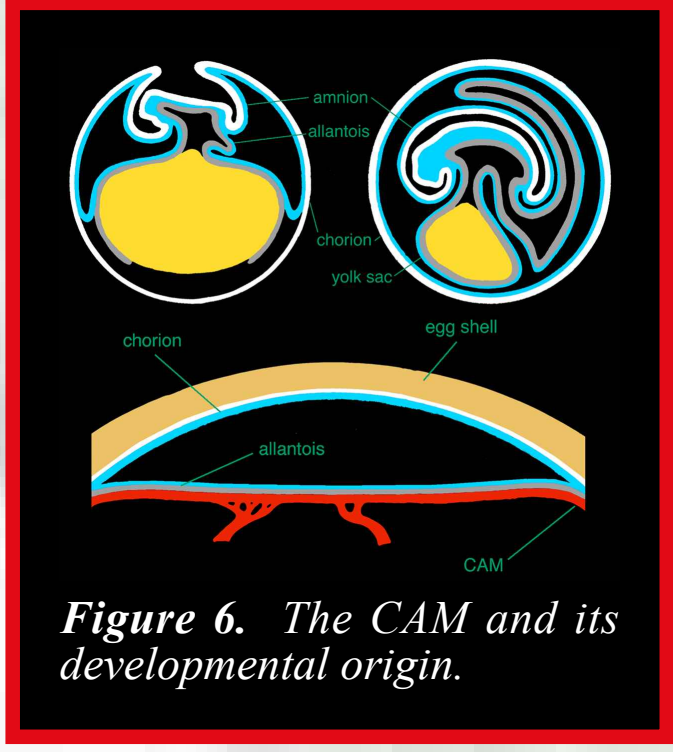


Figure 6. The CAM and its developmental origin.

Structure & Function

**Random mosaic molecules and X-ray crystallography**

The crystal structure of VEGF has been resolved as well as the structure of the VEGF-Flt-1 complex. The interaction between VEGF and KDR has not been resolved at structural level and almost nothing is known about the KDR-VEGF-C interaction. To pinpoint the important elements of receptor specificity we started the determination of VEGF-C structure by X-ray crystallography and the creation of a mammalian expression library of 512 different VEGF-VEGF-C mosaic molecules, whose receptor specificities are correlated to the individual composition of the clone. Key features of this approach are the exploitation of the high homology of VEGF to VEGF-C in the core region to create the library and the use of an efficient screening system, that evaluates the receptor affinities of the individual clones.

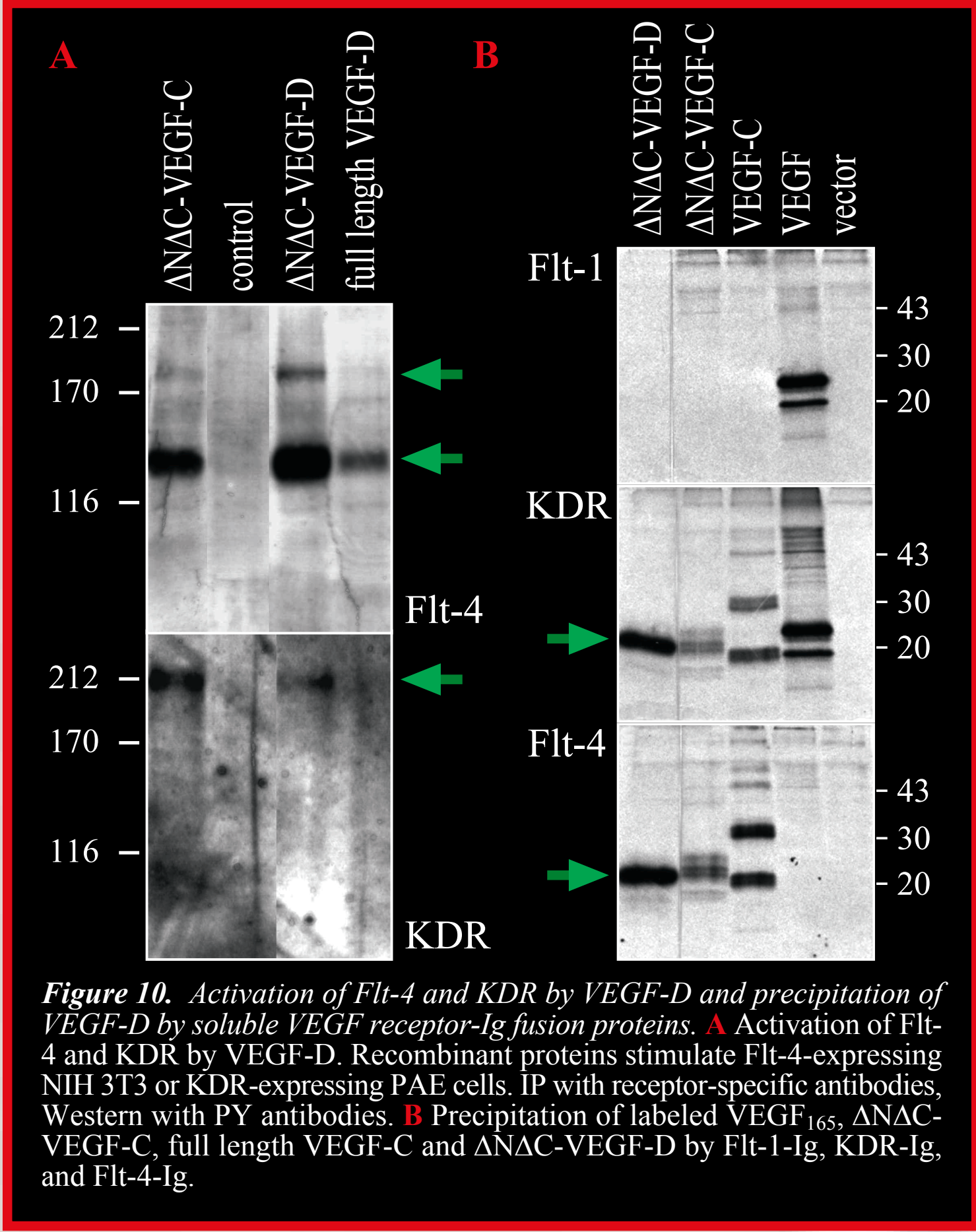


Figure 10. Activation of Flt-4 and KDR by VEGF-D and precipitation of VEGF-D by soluble VEGF receptor-Ig fusion proteins. A: Activation of Flt-4 and KDR by VEGF-D. Recombinant proteins stimulate Flt-4-expressing NIH 3T3 or KDR-expressing PAE cells. IP with receptor-specific antibodies. Western with PY antibodies. B: Precipitation of labeled VEGF<sub>165</sub>, ANAC-VEGF-C, full length VEGF-C and ANAC-VEGF-D by Flt-1-Ig, KDR-Ig, and Flt-4-Ig.

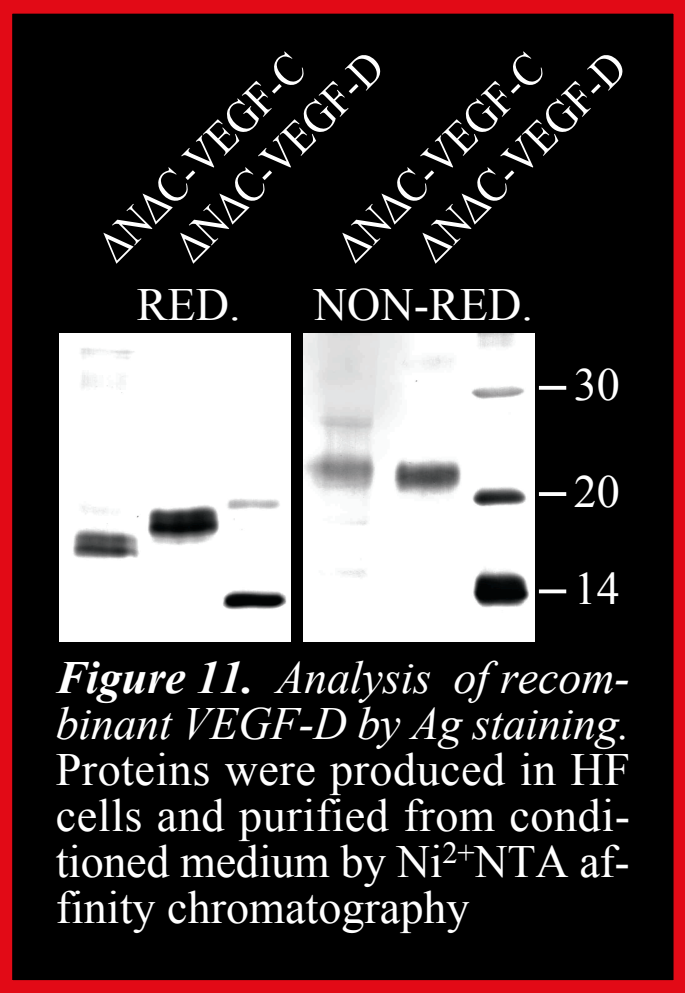


Figure 11. Analysis of recombinant VEGF-D by Ag staining. Proteins were produced in Hi5 cells and purified from conditioned medium by Ni<sup>2+</sup>NTA affinity chromatography

More about VEGF-C & D

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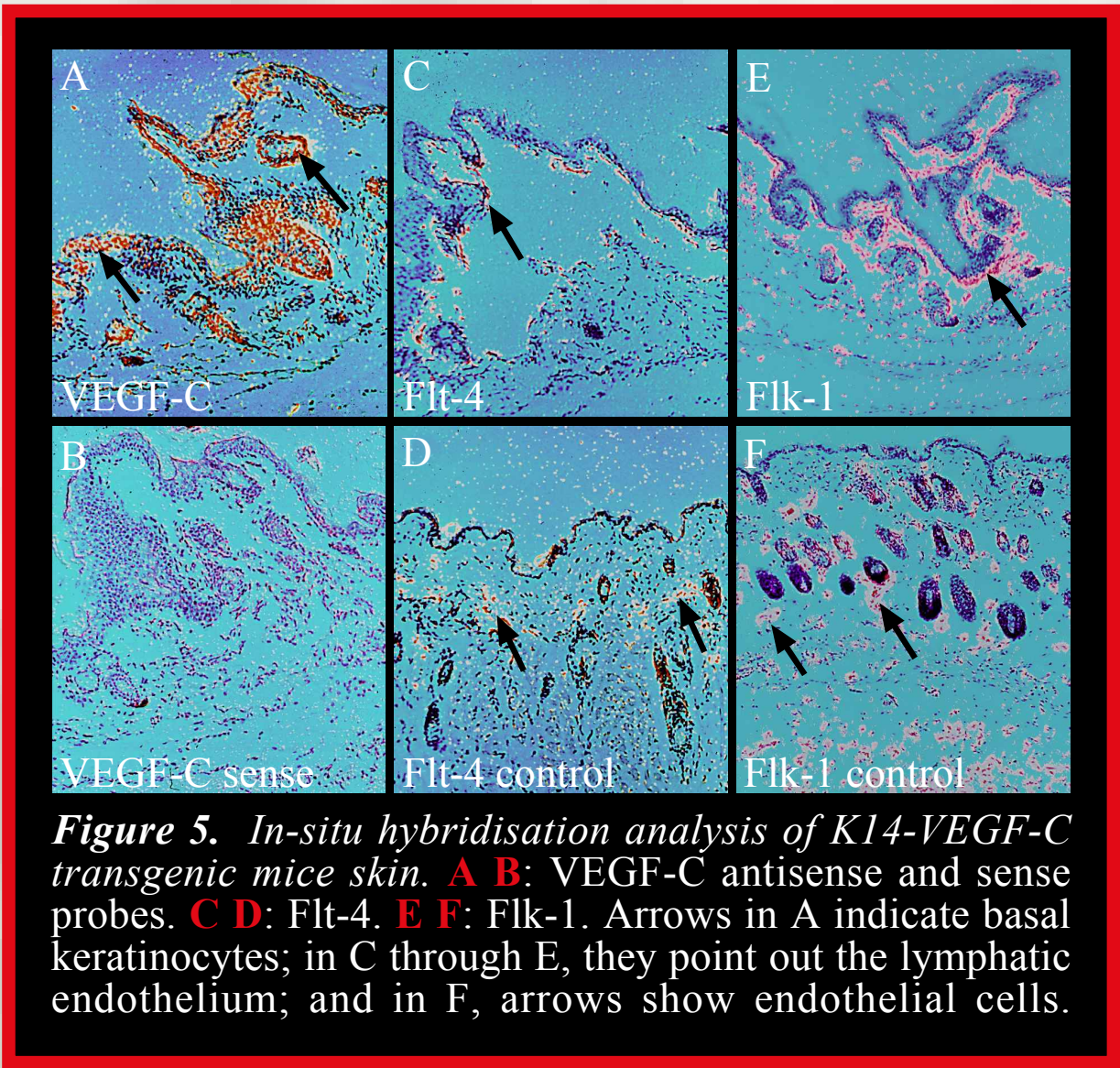


Figure 5. In-situ hybridisation analysis of K14-VEGF-C transgenic mice skin. A: VEGF-C antisense and sense probes. C: D: Flt-4. E: F: Flk-1. Arrows in A indicate basal keratinocytes; in C through E, they point out the lymphatic endothelium; and in F, arrows show endothelial cells.

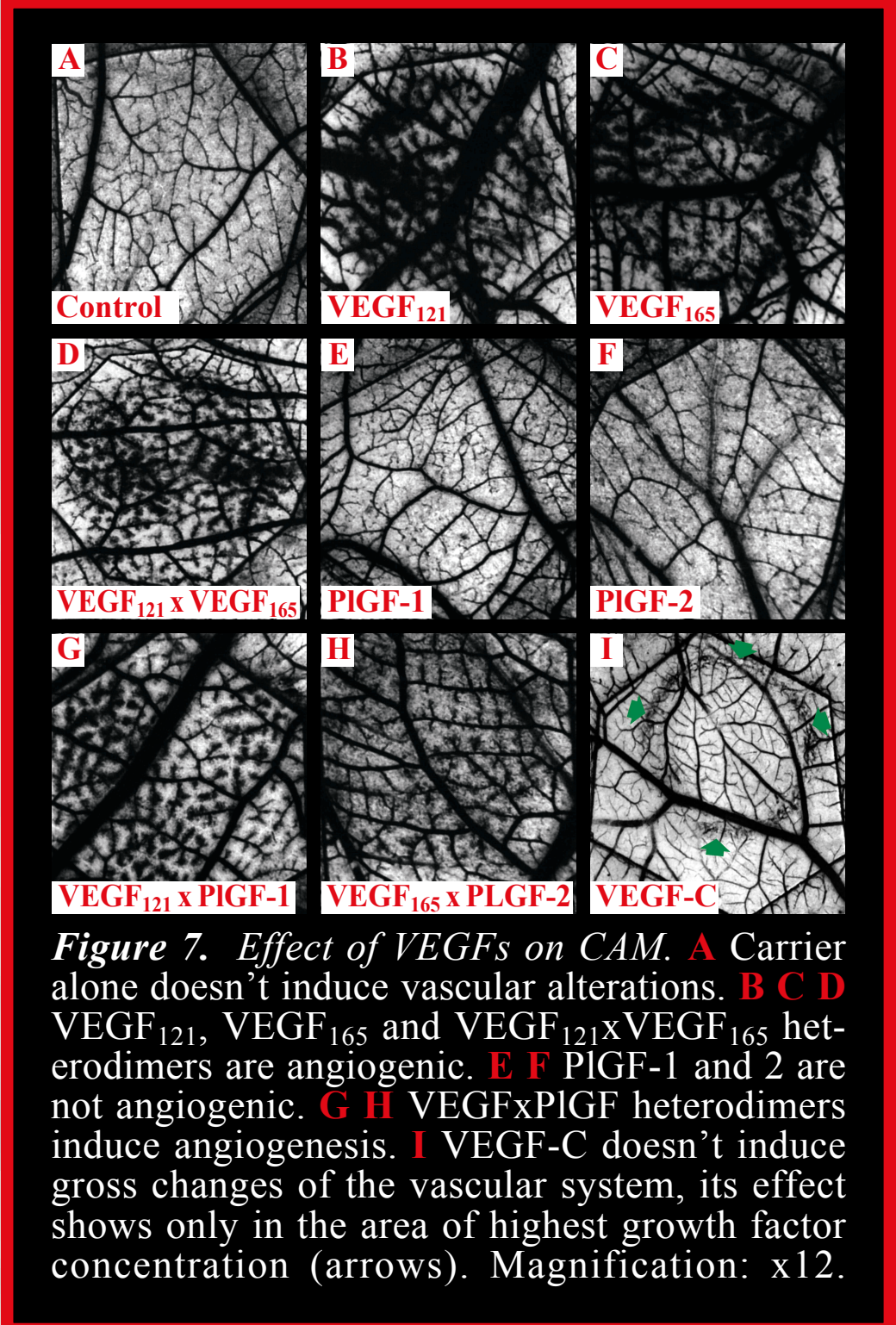


Figure 7. Effect of VEGFs on CAM. A: Carrier alone doesn't induce vascular alterations. B: C: D: VEGF<sub>121</sub>, VEGF<sub>165</sub> and VEGF<sub>121</sub>xVEGF<sub>165</sub> heterodimers are angiogenic. E: F: PlGF-1 and 2 are not angiogenic. G: H: VEGF<sub>x</sub>PlGF heterodimers induce angiogenesis. I: VEGF-C doesn't induce gross changes of the vascular system, its effect shows only in the area of highest growth factor concentration (arrows). Magnification: x12.

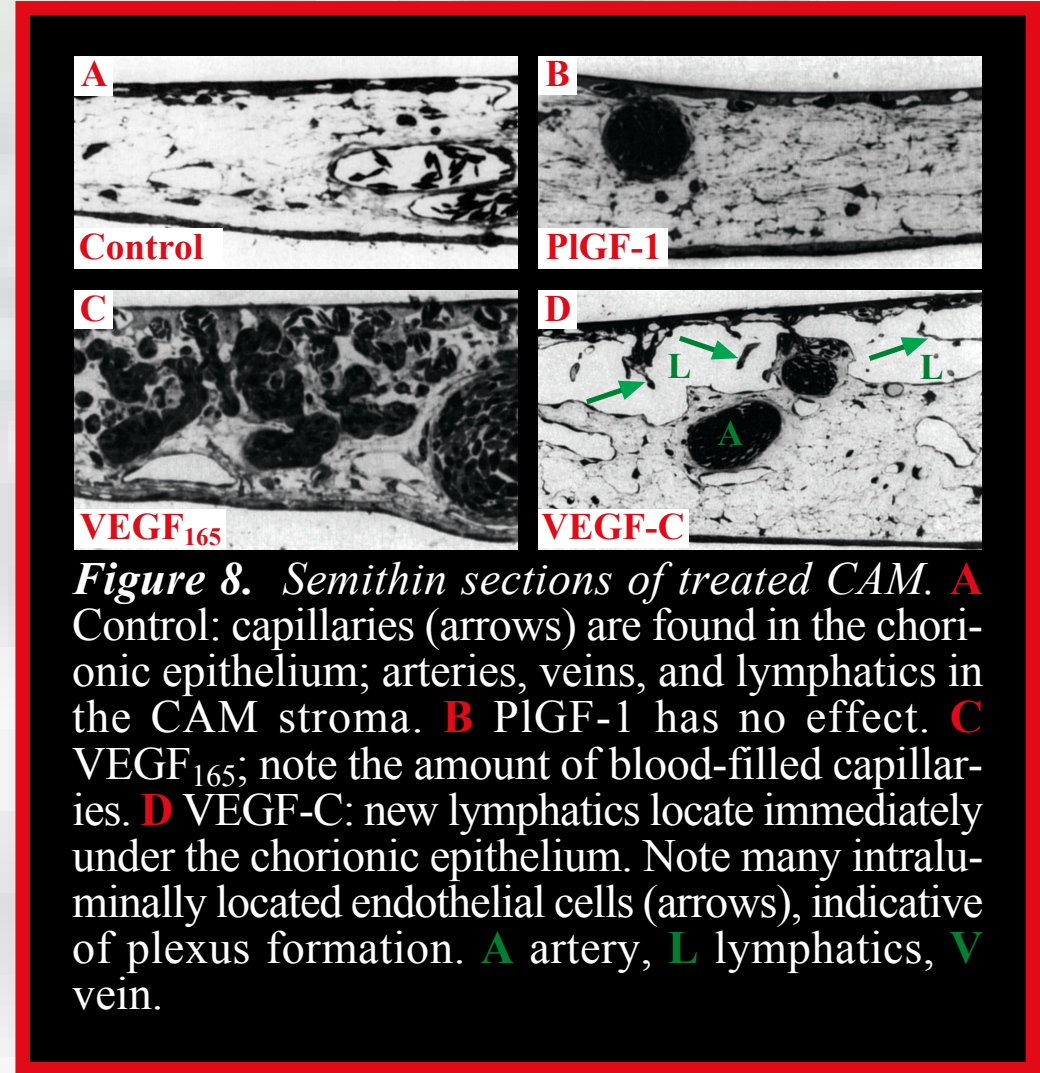


Figure 8. Semithin sections of treated CAM. A: Control. capillaries (arrows) are found in the chorio-epithelium, arteries, veins, and lymphatics in the CAM stroma. B: PlGF-1 has no effect. C: VEGF<sub>165</sub>; note the amount of blood-filled capillaries. D: VEGF-C; new lymphatics locate immediately under the chorio-epithelium. Note many intraluminally located endothelial cells (arrows), indicative of plexus formation. A: artery, L: lymphatics, V: vein.

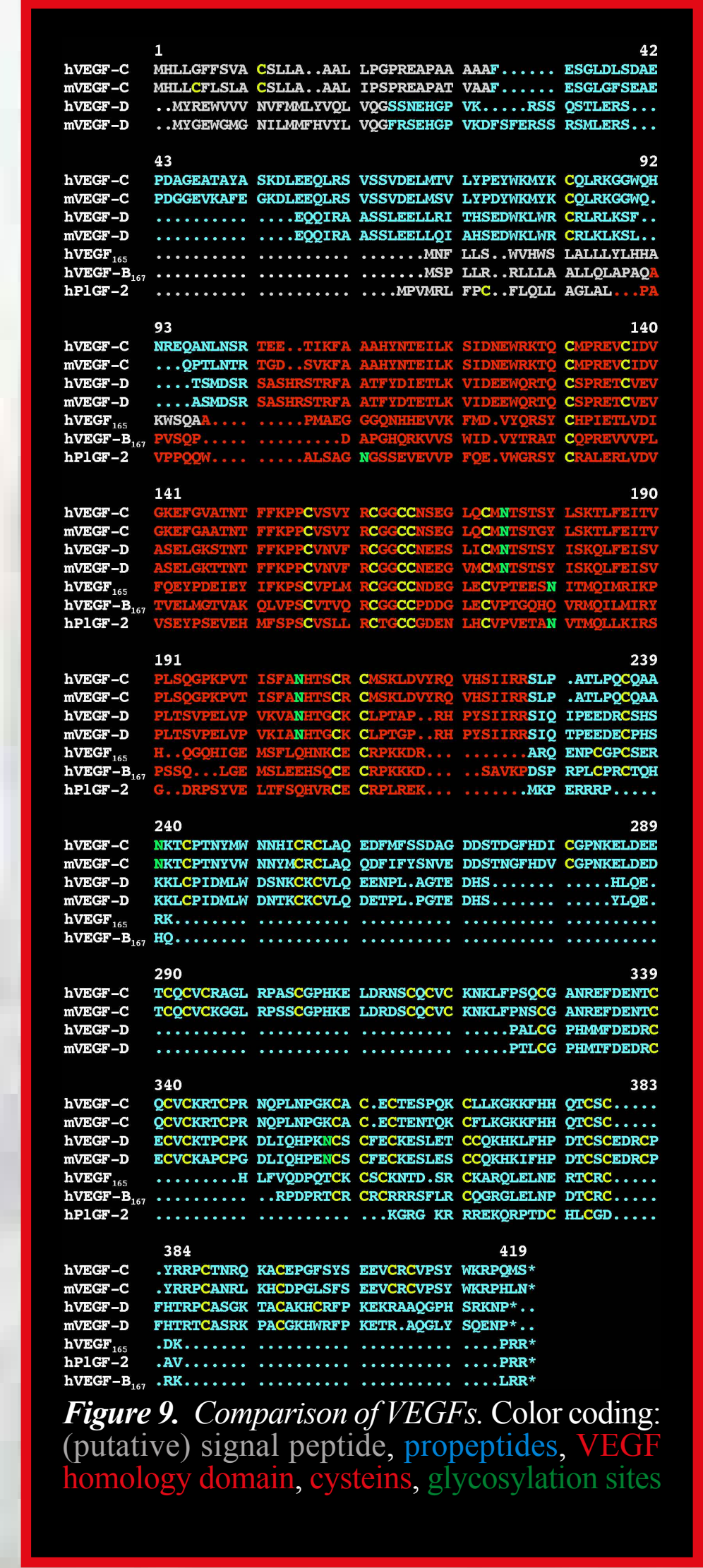


Figure 9. Comparison of VEGFs. Color coding: (putative) signal peptide, propeptides, VEGF homology domain, cysteins, glycosylation sites

The Alphabet of Angiogenesis

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